

Exploiting peptide presentation for the development of novel immunotherapies

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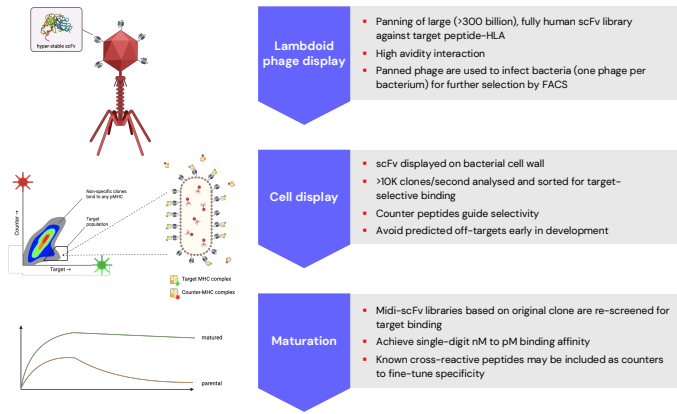
Background

- Most immunotherapies target cell surface proteins preferentially expressed by tumors and not healthy tissue.
 - Many high profile targets, such as common oncogenic driver mutations, are found intracellularly and inaccessible to conventional immunotherapeutic approaches.
 - However, peptides derived from intracellular proteins that are presented on the cell surface as peptide-Human Leukocyte Antigen (HLA) complexes may be targeted.
- Myrio's platform, Retained Display (ReD)TM, enables efficient isolation and affinity maturation of specific peptide-HLA complex binders (single chain variable fragments, scFvs), that are readily converted to different therapeutic formats, including CAR and bispecific antibodies.

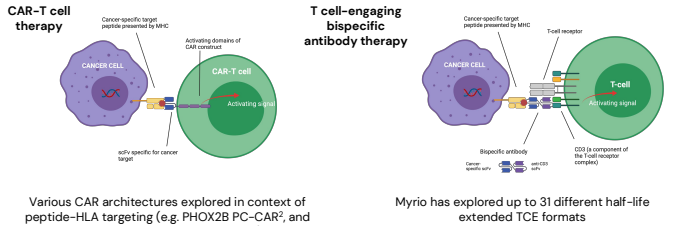
Peptide classes successfully targeted using ReDTM

Neoantigens Driver mutations, E.g. mutant KRAS Gene fusions, E.g. EWS-WT1	Developmental/ cancer testis antigens E.g. PRAME, PHOX2B	Virus-derived E.g. human endogenous retrovirus (HERV-E), human papilloma virus (HPV16 E7)	Secreted tumor- associated factors E.g. Prostate Specific Antigen (PSA)	'Dark antigens' E.g. novel epitopes generated by alternative splicing
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ReDTM platform for peptide-HLA binder discovery and maturation



scFvs can be converted to different therapeutic formats



Example 1: Isolation of a binder 'breaking restriction' across HLA alleles

- Target:** 10-mer peptide derived from human endogenous retrovirus type E (HERV-E) presented by HLA-A*11:01
- Target indication:** clear cell renal carcinoma (ccRCC)
- An scFv isolated by ReDTM** recognizes the same HERV-E peptide presented by HLA-A*11:01 and HLA-A*03:01

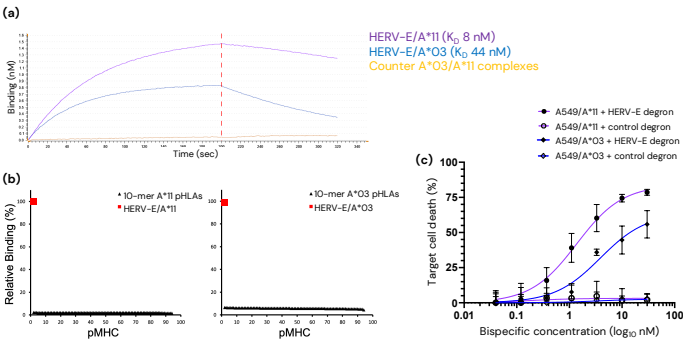


Figure 1: ReDTM scFv library was screened for binders of a 10-mer peptide derived from HERV-E in HLA-A*11. (a) Binding kinetics of an isolated scFv to HERV-E/A*11 (purple) and HERV-E/A*03 (blue), assessed using Biolayer Interferometry. (b) Target selectivity assessed using arrays of irrelevant 10-mer peptides refolded in A*11 (left) or A*03 (right), relative to target HERV-E/A*11 or A*03, using a bead-based flow cytometric assay. (c) scFv was converted to T cell engager format by combining with an anti-human CD3 scFv and evaluated for the ability to mediate T cell killing of luciferase⁺ A549 cells engineered to endogenously present HERV-E peptide by A*11 (closed circles, purple) or A*03 (closed diamond, blue).

References:

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Example 2: Isolation of binders targeting a mutant KRAS neoantigen peptide

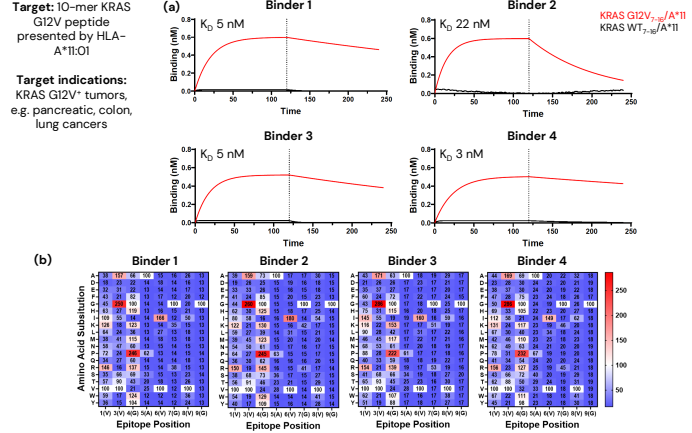


Figure 2: ReDTM scFv library was screened for binders of 10-mer KRAS G12V/A*11, while avoiding wildtype KRAS/A*11. (a) Binding kinetics of isolated scFvs to KRAS G12V/A*11 (red) and wildtype KRAS/A*11 (black) complexes assessed using Biolayer Interferometry. (b) X-Scan analysis was used to map the 'footprint' of each scFv on the target complex. Each position of the target peptide (excluding anchor residues) was substituted for every amino acid (excluding cysteine) and refolded as pHLA. Binding was assessed using a fluorometric bead-based assay, relative to the target complex.

Affinity matured binders maintain target selectivity

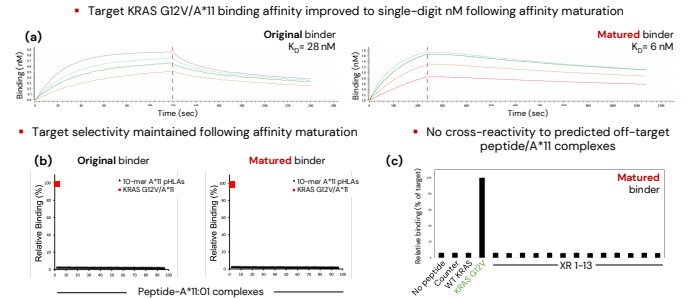


Figure 3: Anti-KRAS G12V/A*11 scFv originally isolated using ReDTM was successfully affinity matured to single-digit nM affinity while maintaining target selectivity. (a) Target binding affinity assessed using different concentrations of KRAS G12V/A*11 complex by Biolayer Interferometry. (b) Target selectivity assessed using an array of 10-mer peptides refolded in A*11 using a bead-based flow cytometric assay. (c) Predicted off-target peptides with homology to KRAS G12V 10-mer peptide were refolded with A*11 and assessed for scFv binding using a bead-based flow cytometric assay.

KRAS G12V/A*11 targeting T cell engager activity in vitro

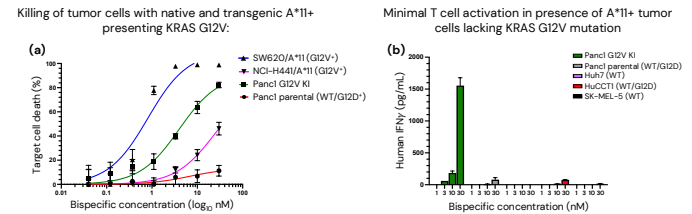


Figure 4: An scFv converted to half-life extended T cell engager (TCE) format was evaluated for activity against KRAS mutant and wild type tumor models. (a) Varying degrees of potency against tumor models presenting different levels of KRAS G12V/A*11 target complex. A Panc1 KRAS G12V knockin (KI) model was generated using CRISPR editing of parental Panc1 (A*11). Killing was assessed by measuring luciferase activity from target cells 48 hours after T cells and antibody were added at 5:1 E:T ratio. (b) Using a panel of A*11+ tumor models, IFN γ release was observed only in the presence of KRAS G12V⁺ cells, but not KRAS wild type of G12D⁺ cells (measured by ELISA).

KRAS G12V/A*11 CAR-T activity in vitro

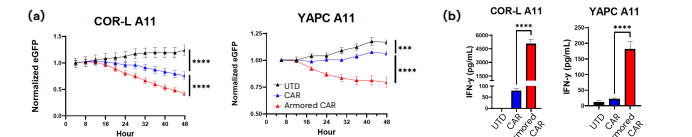


Figure 5: An scFv was converted to CAR and armored CAR formats. Transduced primary T cells were evaluated for activity against KRAS G12V/A*11 tumor model spheroids over 48 hours. (a) Killing of COR-L A*11 and YAPC A*11 spheroids measured using fluorescence microscopy. (b) IFN γ release by CAR- and armored CAR-T measured by ELISA.

Conclusions

- Retained DisplayTM identifies binders of specific peptide-HLA complexes for therapeutic targeting of solid tumors.
- Depending on the desired clinical strategy, scFvs convert easily to different therapeutic formats and can be affinity matured without losing target selectivity.
- scFvs may 'break restriction', recognising the same target peptide presented by different HLA alleles, which significantly expands eligible patient populations.

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